

An Update on the Molecular Alterations and Inflammation Levels of Peri-implant Tissues

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World Journal of Dentistry (2022): 10.5005/jp-journals-10015-2126

In the last years, dental implant therapy became very common, due to the higher quality results and the great predictability of the treatment.¹ Often, despite the current high-level endodontic imaging techniques, in the presence of complex molar or premolars endodontic retreatments, in many cases it is preferred to resort to the implant-prosthetic solution, which is sometimes more predictable in the long term, and often with better esthetic outcomes, and shorter operating times.²⁻⁵

Results of recent studies suggest that soft tissues around implants may be characterized by a higher proinflammatory state compared with soft tissues around teeth, despite adequate implant-prosthetic planning, correct implant emergence, correct prosthetic profiles, and adequate esthetics.^{1,6-9} Controlling and, if possible, reducing the levels of this inflammation can guarantee long-term implant success, and slow down if not completely cancel peri-implant bone loss.^{6,10-12}

Comparing the crevicular fluid molecular composition around dental implants placed with one-stage protocol and surgically accessed adjacent teeth, the authors found higher levels of pro-inflammatory cytokines around implants.^{1,11} Peri-implant higher inflammation levels and early peri-implant bone resorption have been mainly explained by the effect of the inflammatory cell infiltration produced by the implant-abutment interface (micro-gap), and connective tissue stabilization, whereby the circular fibers on the rehabilitation surface protect the internal ones.¹³ Moreover, the presence of cement-retained implant restorations, which may have produced intrasulcular cement remnants, can also cause an increase in peri-implant inflammation levels and subsequently the onset of peri-implantitis phenomena.^{14,15}

Achieving soft tissue stabilization as coronal as possible is one of the main objectives in obtaining esthetic results, and may help to diminish early peri-implant bone resorption.¹⁶ Further evidence in the literature reports how some surfaces and surface treatments of the implant neck guarantee greater adhesion of fibroblasts and therefore better long-term tissue stability.^{1,11,16} When the connective tissue is stabilized, it prevents the apical migration of the epithelium and dictates how much bone resorption occurs.¹⁷

The peri-implant tissues (connective tissue and epithelium) have two functions regarding the protective role. The first one is similar to periodontal sealing and adhesion on the tooth which mitigates bacterial contamination.^{7,8,18,19} The second role is related to mechanical tissue stability around the implant.¹⁸

The supracrestal area: This part of the restoration is a key factor for maintaining the connective tissue at the supracrestal level and, in turn, for bone preservation. It is now clear the importance of the connective tissue around the implants, and of the adequate thickness of the peri-implant tissues, keratinized and not: in this

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How to cite this article: Abbagnale R, Bhandi S, Pagnoni F, et al. An Update on the Molecular Alterations and Inflammation Levels of Peri-implant Tissues. *World J Dent* 2022;13(6):543-544.

Source of support: Nil

Conflict of interest: None

regard, the evidence regarding the grafts of matrices is always greater, with increasingly better esthetic outcomes, and less biological damage for the patient.^{20,21} The abutment, in theory, must fulfill this role.¹² Unfortunately, the implant abutment lacks the capability to retain the connective tissue fibers by means of periodontal fiber insertion, unless the current possibilities of also using prosthetic components that have undergone laser treatments are exploited.^{11,12} This inherent deficiency forces an apical migration of the connective tissue fibers until they are retained at the level of the first thread of the implant.

It is now clear that a correct prosthetic load is essential in the early stages of implant integration, with materials that can absorb part of the occlusal load and reduce stress at the bone-implant level, increasing the predictability of the treatment.²²

REFERENCES

- Guarnieri R, Miccoli G, Reda R, et al. Sulcus fluid volume, IL-6, and IL-1b concentrations in periodontal and peri-implant tissues comparing machined and laser-microtextured collar/abutment surfaces during 12 weeks of healing: a split-mouth RCT. *Clin Oral Implants Res* 2022;33(1):94-104. DOI: 10.1111/clr.13868
- Reda R, Zanza A, Cicconetti A, et al. Ultrasound imaging in dentistry: a literature overview. *J Imaging* 2021;7(11):238. DOI: 10.3390/jimaging7110238
- Di Nardo D, Gambarini G, Capuani S, et al. Nuclear magnetic resonance imaging in endodontics: a review. *J Endod* 2018;44(4):536-542. DOI: 10.1016/j.joen.2018.01.001
- Gambarini G, Plotino G, Grande NM, et al. Differential diagnosis of endodontic-related inferior alveolar nerve paraesthesia with cone beam computed tomography: a case report. *Int Endod J* 2011;44(2):176-181. DOI: 10.1111/j.1365-2591.2010.01816.x
- Reda R, Zanza A, Bhandi S, et al. Surgical-anatomical evaluation of mandibular premolars by CBCT among the Italian population. *Dent Med Probl* 2022;59(2):209-216. DOI: 10.17219/dmp/143546

6. Guarnieri R, Zanza A, D'Angelo M, et al. Correlation between Peri-implant marginal bone loss progression and peri-implant sulcular fluid levels of metalloproteinase-8. *J Pers Med* 2022;12(1):58. DOI: 10.3390/jpm12010058
7. Mahendra J, Mahendra L, Mugri MH, et al. Role of periodontal bacteria, viruses, and placental *mir155* in Chronic periodontitis and preeclampsia—a genetic microbiological study. *Curr Issues Mol Biol* 2021;43(2):831–844. DOI: 10.3390/cimb43020060
8. Balaji TM, Varadarajan S, Jagannathan R, et al. Melatonin as a Topical/systemic formulation for the management of periodontitis: a systematic review. *Materials (Basel)* 2021;14(9):2417. DOI: 10.3390/ma14092417
9. Alhammadi MS, Al-Mashraqi AA, Alnami RH, et al. Accuracy and reproducibility of facial measurements of digital photographs and wrapped cone beam computed tomography (CBCT) photographs. *Diagnostics (Basel)* 2021;11(5):757. DOI: 10.3390/diagnostics11050757
10. Perrotti G, Baccaglione G, Clauser T, et al. Total face approach (TFA) 3D Cephalometry and superimposition in orthognathic surgery: evaluation of the vertical dimensions in a consecutive series. *Methods Protoc* 2021;4(2):36. DOI: 10.3390/mps4020036
11. Guarnieri R, Miccoli G, Reda R, et al. Laser microgrooved vs. machined healing abutment disconnection/reconnection: a comparative clinical, radiographical and biochemical study with split-mouth design. *Int J Implant Dent* 2021;7(1):19. DOI: 10.1186/s40729-021-00301-6
12. Guarnieri R, Reda R, Di Nardo D, et al. Clinical, radiographic, and biochemical evaluation of two-piece versus one-piece single implants with a laser-microgrooved collar surface after 5 years of functional loading. *Clin Implant Dent Relat Res* 2022. DOI: 10.1111/cid.13118
13. Ericsson I, Persson LG, Berglundh T, et al. Different types of inflammatory reactions in peri-implant soft tissues. *J Clin Periodontol* 1995;22(3):255–261. DOI: 10.1111/j.1600-051x.1995.tb00143.x
14. Berglundh T, Lindhe J. Dimension of the periimplant mucosa. Biological width revisited. *J Clin Periodontol* 1996;23(10):971–973. DOI: 10.1111/j.1600-051x.1996.tb00520.x
15. Reda R, Zanza A, Cicconetti A, et al. A Systematic review of cementation techniques to minimize cement excess in cement-retained implant restorations. *Methods Protoc* 2022;5(1):9. DOI: 10.3390/mps5010009
16. Rodríguez X, Vela X, Calvo-Guirado JL, Nart J, Stappert CF. Effect of platform switching on collagen fiber orientation and bone resorption around dental implants: a preliminary histologic animal study. *Int J Oral Maxillofac Implants* 2012;27(5):1116–1122.
17. Rompen E, Domken O, Degidi M, et al. The effect of material characteristics, of surface topography and of implant components and connections on soft tissue integration: a literature review. *Clin Oral Implants Res* 2006;17 Suppl 2:55–67. DOI: 10.1111/j.1600-0501.2006.01367.x
18. Marconcini S, Giammarinaro E, Covani U, et al. The effect of tapered abutments on marginal bone level: a retrospective cohort study. *J Clin Med* 2019;8(9):1305. DOI: 10.3390/jcm8091305
19. Flamm J, Hartung S, Gänger S, et al. Establishment of an olfactory region-specific intranasal delivery technique in mice to target the central nervous system. *Front Pharmacol* 2021;12:789780. DOI: 10.3390/coatings11091035
20. Guarnieri R, Reda R, Di Nardo D, et al. In vitro direct and indirect cytotoxicity comparative analysis of one pre-hydrated versus one dried acellular porcine dermal matrix. *Materials (Basel)* 2022;15(5):1937. DOI: 10.3390/ma15051937
21. Tavelli L, Barootchi S, Avila-Ortiz G, et al. Peri-implant soft tissue phenotype modification and its impact on peri-implant health: a systematic review and network meta-analysis. *J Periodontol* 2021;92(1):21–44. DOI: 10.1002/JPER.19-0716
22. Katheng A, Kanazawa M, Komagamine Y, et al. Masticatory performances and maximum occlusal forces of immediate and conventional loaded two-implant supported overdentures retained by magnetic attachments: preliminary study of randomized controlled clinical trial. *Int J Implant Dent* 2021;7(1):57. DOI: 10.1186/s40729-021-00342-x